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AUTHORS: Asiye BERBER,İsmühan POTOGLU ERKARA,Murat OLGUN,Onur KOYUNCU,Murat ARDIÇ,Okan SEZER

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Heavy metal content screening in leaves and flowers of *Hypericum organifolium* by atomic absorption spectrometry

Asiye BERBER¹, İsmühan POTOĞLU ERKARA^{*2}, Murat OLGUN², Onur KOYUNCU², Murat ARDIÇ²,
Okan SEZER²

ORCID: 0000-0002-8340-4793; 0000-0001-5780-4999; 0000-0002-6234-455X; 0000-0002-0364-6638;
0000-0001-8734-3038; 0000-0001-7304-1346

¹ Eskişehir Osmangazi Univ, Faculty of Education, Department of Elementary Science Education, Eskişehir, Turkey

² Eskişehir Osmangazi University, Faculty of Science and Letters, Department of Biology, Eskişehir, Turkey

³ Eskişehir Osmangazi University, Agricultural Faculty, Field Crop Department, Eskişehir, Turkey

Abstract

Atomic absorption spectrometry facilitates the reliable determination of mineral content during pharmaceutical quality control of medicinal plants. In the present work, measurable amounts of Fe, Ca, Cu, K, Mg, Mn, Na and Zn were detected in the leaves and flowers of *Hypericum organifolium* Willd. through atomic absorption spectrometry. Mean heavy metal content in the flowers and leaves of *H. organifolium* was, in descending order, Ca > Mg > K > Na > Fe. Ca was present in higher concentrations in the flowers (11157.24 ppm) and leaves (20132.24 ppm) of titled plant. Our results reveal that flowers are less suitable as target plant parts for metal accumulation than leaves.

Keywords: *Hypericum organifolium*, heavy metals, atomic absorption spectrometry

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Hypericum organifolium'un yaprak ve çiçeklerindeki ağır metal içeriğinin atomik absorpsiyon spektrometresi ile belirlenmesi

Özet

Atomik absorpsiyon spektrometresi, tıbbi bitkilerin farmasötik kalite kontrolü sırasında mineral içeriğinin güvenilir bir şekilde belirlenmesini kolaylaştırmaktadır. Bu çalışmada, *Hypericum organifolium* Willd'un yaprak ve çiçeklerinde atomik absorpsiyon spektrometresi ile ölçülebilir miktarda Fe, Ca, Cu, K, Mg, Mn, Na ve Zn bulunduğu tespit edilmiştir. *H. organifolium*'un yaprak ve çiçeklerindeki ortalama ağır metal içeriği sırasıyla Ca > Mg > K > Na > Fe olarak belirlenmiştir. Kalsiyum, çiçek (11157,24 ppm) ve yapraklarda (20132,24 ppm) en yüksek konsantrasyonda tespit edilmiştir. Elde edilen sonuçlar çiçeklerin yapraklara kıyasla çok daha düşük seviyede ağır metal birikimi gösterdiğini ortaya koymaktadır.

Anahtar kelimeler: *Hypericum organifolium*, ağır metaller, atomik absorpsiyon spektrometresi

1. Introduction

Especially in developing countries, environmental pollutants which are released to nature in an uncontrolled manner reach the structure of plants through water, soil and air. Metals have an important place among these pollutants. The use of metal based pollutant exposed plants by humans causes serious health problems such as kidney damage, renal failure and liver damage [1; 2; 3]. Iron, zinc, calcium, magnesium, copper, potassium, manganese, sodium were chosen as representative metals whose levels in the environment represent a reliable index of environmental pollution and human health. Some plants like as *Hypericum perforatum*, *Avena sterilis*, *Bifora radians*, *Chenopodium album*,

* Corresponding author / Haberleşmeden sorumlu yazar: Tel.: +905326338464; Fax.: +905326338464; E-mail: ismuhan@ogu.edu.tr

Consolida regalis, *Humulus lupulus*, *Reseda lutea*, *Solanum nigrum*, *Sorghum halepense* and *Xanthium strumarium* can then be used as biomonitors for the determination of trace element levels [4; 5; 6; 7].

Hypericum L., which is a member of the Hypericaceae (Guttiferae) family, is represented approximately 500 species all around to world. About 108 of these species are naturally distributed in Turkey [8; 9; 10; 11]. There has been many studies previously conducted on the *Hypericum* species, but very limited studies for *H. origanifolium*. According to the literature reviews, only one anatomical study on *H. origanifolium* was found [12]. One of the most important features of *Hypericum* taxa is their unique secondary metabolites. *Hypericum* taxa and their metabolites are widely used in traditional and modern medicine today [13]. In the light of phytochemical studies performed on *H. origanum*, flavonoids (myrcetin, rutin, quercetin, hyperoside), xanthenes (iso-magniferin and mangiferin), naphthodiantrones (frangulin, emodin, proto-pseudohypericin, pseudohypericin and phenolicin) and phenolic acids have been determined at the aerial parts of the plant [14; 15; 16; 17; 18; 19; 20].

Data available on the biological activity of *H. origanifolium* is also limited to a few previous reports that have demonstrated their cytotoxic, antiproliferative, antimicrobial, antibacterial, antiyeast and antioxidant activities [18; 19; 20; 21; 22; 23; 24; 25; 26]. In the light of literature data, any studies on the heavy metal content of *H. origanifolium* not determined.

The main purpose of this study is to determine the heavy metal content in extracts obtained from the flowers and leaves of *H. origanifolium* by atomic absorption spectrometry. We hope that the relationship between the heavy metal amounts in *H. origanifolium* leaves and flowers will be reference for future studies.

2. Materials and methods

H. origanifolium Willd. was collected around of Eskişehir, Turkey. *H. origanifolium* Willd. Sivrihisar, Tekören village, 1100 m. June 2003 (OUFE 10334). The plant was identified according to Flora of Turkey and the East Aegan Islands [8; 27; 28].

For the solid samples with a nitric acid (Merck, Darmstadt, Germany)–perchloric acid (Merck, Darmstadt, Germany) digestion was used for mineralizing. The dried flowers and leaves of *H. origanifolium* were extracted for the solution phase as described previously and analyzed for Fe, Zn, Ca, Mg, Cu, K, Mn, Na (Merck AAS standard solutions), using Hitachi (180-70) Polarized Zeeman flame atomic absorption spectrometry [29; 30]. All precautions were taken to prevent metal contamination, i.e. samples were cleaned with 2% HNO₃, rinsed in distilled water and baked at 600°C. All samples were analyzed in triplicate and the mean values were calculated. In order to increase the reliability of the measurements during the study, the instrument was calibrated at every 10 readings.

The flame atomic absorption spectrometry (FAAS) instrumental and operating conditions that provided the best sensitivity for the determination of metal content are detailed in Table 2.1.

Table 2.1. FAAS instrumental parameters employed to determine metals

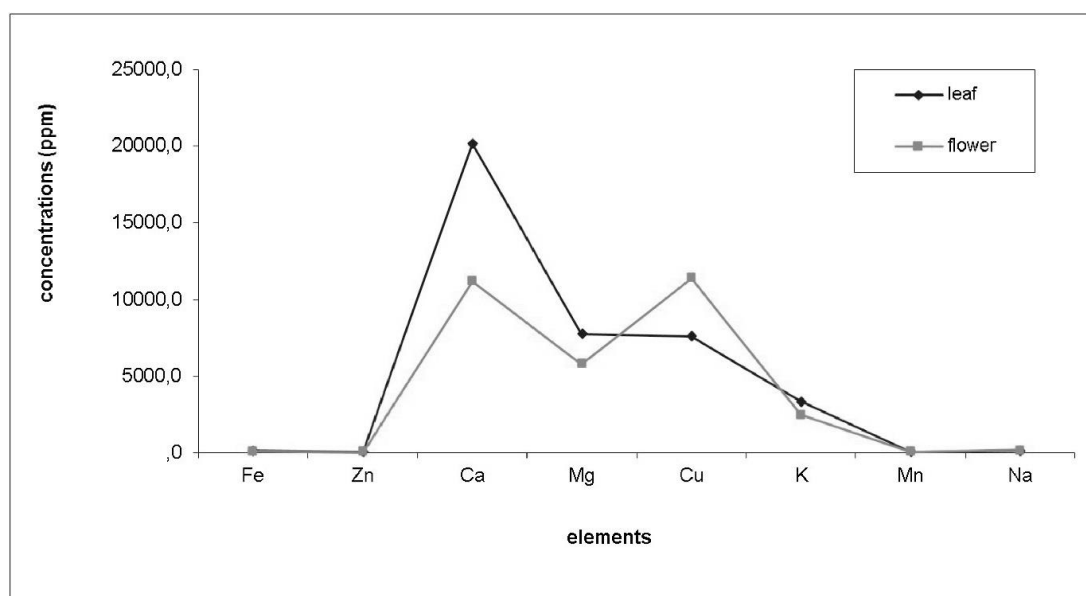
Elements	Flame type	Burner height (mm)	Wavelength (nm)	Slit width (nm)	Lamp Current (mA)	Fuel gas (1 min ⁻¹)
Fe	Air-C ₂ H ₂	7.5	248.3	0.2	10	2.3
Zn	Air-C ₂ H ₂	7.5	213.8	1.3	10	2.0
Ca	Air-C ₂ H ₂	12.5	422.7	2.6	7.5	2.6
Mg	Air-C ₂ H ₂	7.5	285.2	2.6	7.5	1.6
Cu	Air-C ₂ H ₂	7.5	324.8	1.3	7.5	2.3
K	Air-C ₂ H ₂	7.5	766.5	2.6	10	2.3
Mn	Air-C ₂ H ₂	7.5	279.5	0.4	7.5	2.3
Na	Air-C ₂ H ₂	7.5	589.0	0.4	10	2.2

3. Results

In this study, the heavy metal contents of the leaves and flowers of *Hypericum origanifolium* Willd. were investigated. These levels were obtained through flame atomic absorption spectrometry. Fe, Zn, Ca, Mg, Cu, K, Mn and Na were determined to be present in the samples. Metal concentrations in the leaves of *H. origanifolium* were found to be 130.25, 30.99, 20132.24, 7775.84, 7.60, 3361.53, 48.40, and 145.02 ppm; and in flower of *H. origanifolium* were found to be 93.64, 26.26, 11157.23, 5768.23, 11.37, 2454.99, 36.12, and 184.94 ppm for iron, zinc, calcium, magnesium, copper, potassium, manganese, sodium, respectively (Table 3.1. and Figure 3.1.).

Table 3.1. Element concentrations of *H. origanifolium* (ppm)

Elements	<i>H. origanifolium</i> (leaf)	<i>H. origanifolium</i> (flower)
Fe	130.25	93.64
Zn	30.99	26.26
Ca	20132.24	11157.23
Mg	7775.84	5768.23
Cu	7.60	11.37
K	3361.53	2454.99
Mn	48.40	36.12
Na	145.02	184.94

Figure 3.1. Relationship between leaf and flower metal concentrations of *H. origanifolium*

4. Conclusions and discussion

In the light of obtained data from flame atomic absorption spectrometry analysis of the leaves and flowers of *Hypericum origanifolium*, Fe, Zn, Ca, Mg, K and Mn concentrations were recorded as higher in the leaf than in the flower, 130.25 > 93.64 ppm, 30.99 > 26.26 ppm, 20132.24 > 11157.23 ppm, 7775.84 > 5768.23 ppm, 3361.53 > 2454.99 ppm, 48.40 > 36.12 ppm, respectively. Cu and Na concentrations were observed to be higher in the flower than in the leaf 11.37 > 7.60 ppm, 184.94 > 145.02 ppm, respectively (Figure 1, Table 2).

The mean heavy metal content in the flowers and leaves of *H. origanifolium* was, in descending order, Ca > Mg > K > Na > Fe. Ca was present in higher concentrations in the flowers (11157.24 ppm) and leaves (20132.24 ppm) of the titled plant. Gomez *et al.* (2004) indicated that the CA concentration in *H. perforatum* was 100-500 ppm for the dried herb, as indicated in table 3.1. [4]. This situation shows that the Ca concentration of the leaf and flower of *H. origanifolium* is higher than *H. perforatum*. Kadioğlu *et al.* found the concentrations to be 495, 62.6, 11.1, 19.5 ppm for iron, manganese, copper, zinc, respectively [5]. In this study, the concentrations of Fe and Mn were higher, while that of Zn was lower than in the *H. perforatum* concentrations reported by Kadioğlu *et al.* (2005).

Findings obtained from elemental analysis studies on plants reveal that herbal foods are rich in especially Mg, Ca, Na and K [3]. The data obtained in our study coincide with the above findings. K, Mg and Ca which are represent the most abundant metal constituents of many plants were identified as the most abundant metals in *H. origanifolium* extracts. Internal and external factors have direct and indirect effects on the different metal concentrations of distinct parts of plants. These regional differences in metal content can occur due to many different reasons, such as genetic factors, growth conditions, analytical procedures and geographical variations. These factors are directly affected by the location differences where the plant samples are collected. Hence, there is an indirect relationship between sample locations and metal content can be mentioned. Though much is known about the functional role of a number of elements, the best foreseeable benefit for human health, mineral nutrition, lies in obtaining the correct amount of supplementation in the right form at the right time. High or low levels of Ca, Cu, Zn, Mn, K and Mg may invite many disorders [3]. These elements also play a part in neurochemical transmission, as well as serving as constituents of biological molecules, as a cofactor for various enzymes such as NAD(P)H oxidase, Ferroxidase, Alcohol dehydrogenase and in a variety of different metabolic processes.

High amounts of Ca are expected one way or another, as it is one of the most common minerals of the soil, from where it is readily absorbed into the plants. Iron is one of the important elements for the human body and is involved in vital activities such as hemoglobin formation, oxygen and electron transfer. Zinc and Copper, on the other hand, are involved in many different vital processes such as being important components of enzymatic and redox systems. For this reason, they have an important place among the elements that should be found regularly in the diet [6]. Also, the consequences show that most of these herbal plants contain vital elements for human metabolism and are also essential for growth and development as well as for the prevention and cure of diseases.

While many investigations into the quality values of medicinal plants are being reported in the current literature [7], less emphasis has been made on the metal content of herbal products. Metallic elements are constituent plant compounds demonstrating biological activity as essential or toxic agents in metabolism. Thus, the application of metal monitoring as a pattern recognition method in medicinal herbs is a promising tool for their characterization.

Chemotaxonomy is one of the most important tools that help classify plant taxa. The field of study of chemotaxonomy is to reveal the chemical content similarities and differences between different taxonomic groups. In this context, determining the metal content of plants is extremely valuable for chemotaxonomic researches. Heavy metal levels are important pollutants for soil, water, plant, the environment and human health. Especially the accumulation of heavy metals in some plant taxa makes them a valuable tool in determining heavy metal pollution. Therefore, further investigations are also needed to determine interactions between the leaves and flowers in terms of heavy metals.

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References

- [1] [Shaw, D., Leon, C., Kolev, S. & Murray, V. (1997). Traditional remedies and food supplements. A 5-year toxicological study (1991-1995). *Drug Safety*, 17, 342-356.
- [2] Andrew, A. S., Warren, A. J., Barchowsky, A., Temple, K. A., Klei, L., Soucy, N. V., O'Hara, K. A. & Hamilton, J. W. (2003). Genomic and Proteomic Profiling of Responses to Toxic Metals in Human Lung Cells. *Environ. Health Perspect*, 111(6), 825-835.
- [3] Ajasa, A. M. O., Bello, M. O., Ibrahim, A. O., Ogunwande, I. A. & Olawore, N. O. (2004). Heavy trace metals and macronutrients status in herbal plants of Nigeria. *Food Chemistry*, 85, 67-71.
- [4] Gomez, M. R., Cerutti, S., Olsina, R. A., Silva, M. F. & Martinez, L. D. (2004). Metal content monitoring in *Hypericum perforatum* pharmaceutical derivatives by atomic absorption and emission spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*, 34(3), 569-576.
- [5] Kadioğlu, I., Mendil, D., Sarı, H. & Hasdemir, E. (2005). Determination of Heavy Metal Levels in Some Weeds Collected from Tokat, Turkey. *Asian Journal of Chemistry*, 17(1), 564-568.
- [6] Özcan, M. M. & Akbulut, M. (2007). Estimation of minerals, nitrate and nitrite contents of medicinal and aromatic plants used as spices, condiments and herbal tea. *Food Chemistry*, 106, 852-858.
- [7] Gomez, M. R., Cerutti, S., Sombra, L. L., Silva, M. F. & Martinez, L. D. (2007). Determination of heavy metals for the quality control in argentinian herbal medicines by ETAAS and ICP-OES. *Food and Chemical Toxicology*, 45, 1060-1064.
- [8] Robson, N. K. B. (1967). *Hypericum* L. – In *Flora of Turkey and the East Aegan Islands* (Ed. Davis PH) Vol. 2. Edinburgh University Press, p. 355.
- [9] Robson, N. K. B. (1977). Studies in the genus *Hypericum* L. (Guttiferae): 1. Infrageneric classification. *Bulletin of the British Museum (Natural History) Botany*, 5: 291-355.
- [10] Robson, N. K. B. (2012). Studies in the genus *Hypericum* L. (Hypericaceae) 9. Addenda, corrigenda, keys, lists and general discussion. *Phytotaxa* 72, 1-111.
- [11] Duman, H. & Çakır-Dindar, E. G. (2020). *Hypericum alacamdaglariense* (Hypericaceae), a new species from Turkey. *Phytotaxa* 470(2), 176-185.
- [12] Potoğlu Erkara, İ. & Tokur, S. (2004). Morphological and Anatomical investigations on some *Hypericum* L., species growing naturally in and around Eskişehir. *Trakya Univ J Sci* 5, 97-105.
- [13] Yaylacı, Ö. K., Özgişi, K., Sezer, O., Orhanoğlu, G., Öztürk, D. & Koyuncu, O. (2013). Anatomical studies and conservation status of rare endemic *Hypericum sechmenii* Ocak & Koyuncu (Sect: Adenosepalum) from Eskişehir-Turkey. *Journal of Selcuk University Natural and Applied Science*, 2(1), 1-11.
- [14] Mathis, C. & Ourisson, G. (1964). Chemo-taxonomic study of the genus *Hypericum*. III. The distribution of saturated hydrocarbons and monoterpenes from the essential oil of *Hypericum*. *Phytochem*, 3, 133-141.
- [15] Kitanov, G. M. & Nedialkov, P. T. (1998). Mangiferin and isomangiferin in some *Hypericum* species. *Biochem Syst Ecol*, 26, 647-653.

- [16] Makovests'ka, O. Y. (1999). Research of Biologically Active Substances of *Hypericum* L. species. Report VI. Sections olympia (Spach) Nyman, oampylopus Boiss. and organifolia Stef. *Farm Zh (Kiev)*, 6, 46-50.
- [17] Sirvent, T. M., Walker, L., Vance, N. & Gibson, D. M. (2002). Variation in hypericins from wild populations of *Hypericum perforatum* L. in the Pacific Northwest of the USA. *Econ Bot*, 56, 41-48.
- [18] Çırak, C., Radušienė, J., Ivanauskas, L. & Janulis, V. (2007). Variation of bioactive secondary metabolites in *Hypericum organifolium* during its phenological cycle. *Acta Physiologiae Plantarum*, 29(3), 197-203.
- [19] Öztürk, N., Tunçel, M. & Potoğlu Erkara, İ. (2009). Phenolic compounds and antioxidant activities of some *Hypericum* species: A comparative study with *H. perforatum*. *Pharmaceutical biology*, 47(2), 120-127.
- [20] Bertoli, A., Çırak, C. & Seyis, F. (2015). *Hypericum organifolium* Willd.: The essential oil composition of a new valuable species. *Industrial Crops and Products*, 77, 676-679.
- [21] Sakar, M. K., Tamer, A. U. & Tokur, S. (1988). Antimicrobial activities of some *Hypericum* species growing in Turkey. *Fitoterapia*, 59, 49-52.
- [22] Sakar, M. K. & Tamer, A. U. (1990). Antimicrobial activity of different extracts from some *Hypericum* species. *Fitoterapia*, 61, 464-466.
- [23] Güzey, G. (2007). Cytotoxic and Antiproliferative Effects of *Hypericum perforatum*, *Hypericum montbretti* and *Hypericum organifolium* Species on A549, HeLa and NIH3T3 Cell Cultures. PhD Thesis, Anadolu University Institute of Health Sciences, Eskişehir, Turkey.
- [24] Güzey, G., Ibadova, S., Öztürk, Y., Öztürk, N., Maggi, F., Sagratini, G., Ricciutelli, M. & Vittori, S. (2011). Antiproliferative and Antioxidant Effects of Three *Hypericum* Species of Turkish Origin: *H. perforatum*, *H. montbretii* and *H. organifolium*. *Medicinal and Aromatic Plant Science and Biotechnology*, 5(1), 91-99.
- [25] Yaşar, Ş. N., Can, Ö. D., Öztürk, N., Sagratini, G., Ricciutelli, M., Vittori, S. & Maggi, F. (2013). Central nervous system activities of *Hypericum organifolium* extract via GABAergic and opioidergic mechanisms. *Phytotherapy Research*, 27(6), 877-884.
- [26] Boran, R. (2018). Investigations of anti-aging potential of *Hypericum organifolium* Willd. for skincare formulations. *Industrial Crops and Products*, 118, 290-295.
- [27] Robson, N. K. B. (1988). *Hypericum* L. – In Flora of Turkey and the East Aegan Islands (Eds. Davis PH, Mill RR and Tan K) Vol.10. Edinburgh University Press, p. 96.
- [28] Dönmez, A. A. (2000). *Hypericum* L. – In Flora of Turkey and the East Aegan Islands (Eds. Güner A, Özhatay N, Ekim T and Başer KHC) Vol.11. Edinburgh University Press, pp. 71-72.
- [29] Que Hee, S. S. & Boyle, Jr. (1988). Simultaneous Multielemental Analysis of Some Environmental And Biological Samples By Inductively Coupled Plasma Atomic Emission-Spectrometry. *Analytical Chemistry*, 60(10), 1033-1042.
- [30] Kılıç, M., Ay, G., Koçbaş, F., Kılıç, F.M. (2019). Determination level of heavy metal in Ayvalik Saltern using *Halimione portulacoides* (L.) plant. *Biological Diversity and Conservation*, 12(1), 100-106.