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Optimizing embryo stage and GA₃ doses in Common mandarin x Carrizo citrange crosses on embryo rescue technique

Yerli mandarin x Carrizo sitranjı melezlerinde, embriyo kurtarma tekniğinde embriyo gelişim aşaması ve GA₃ dozlarının optimizasyonu

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ABSTRACT

Many difficulties exist in citrus breeding due to incompatibility, long juvenility, sterility and nucellar embryo. Zygotic embryo in the seeds of polyembryonic citrus cultivar is degenerated by nucellar embryos during embryo development. Accordingly, in vitro embryo culture is a useful tool in citrus breeding, since it assures embryo germination and development. In this study, Common mandarin x Carrizo citrange were crossed in order to produce new rootstock genotypes. Immature embryos were taken from fruit after 80, 100 and 120 pollination days to determine the suitable embryo rescue stage. Then, the embryos were germinated in Murashige and Tucker (MT) culture medium including 0, 0.5 and 1 mg l⁻¹ GA₃. According to the results, high germination rates were materialized as 100% and on 95% to taken embryos from 120 days after pollination (DAP) in supplemented with 1 mg l⁻¹ GA₃ and 0.5 mg l⁻¹ GA₃ in MT culture medium respectively. The embryos taken from 80 days after pollination did not germinate on MT medium without GA₃ (control) and including with 0.5 mg l⁻¹. Generally, the existence of GA₃ on medium increased the ratio of germination compared to the control. The highest trifoliate ratio was determined as 33.33% at the embryos taken from 100 and 120 days after pollination and medium including 0.5 and 1 mg l⁻¹ GA₃ consecutively. The results have revealed that the optimal time for embryo rescue at citrus is 120 DAP and including GA₃ in the medium increased embryo germination.

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ÖZ

Turunçgil ıslahında; tür ve çeşitler arasında uyumsuzluk, uzun gençlik kısırlığı, gametik kısırlık ve polyembriyoni gibi nedenlere bağlı olarak birçok engel bulunmaktadır. Poliembriyonik turunçgillerin tohumlarındaki zigotik embriyo gelişim sırasında nüseller embriyolar tarafından dejenere edilmektedir. Buna göre, zigotik embriyoların çimlenmesini ve gelişimini sağlaması nedeniyle embriyo kültürü ıslah çalışmalarında yararlı bir araçtır. Bu çalışmada, yeni anaç bireylerinin elde edilmesi amacıyla Yerli mandarin x Carrizo sitranjı melezlenmiştir. Embriyo kurtarma için uygun embriyo gelişim aşamasının belirlenmesi amacıyla olgunlaşmamış embriyolar tozlanmadan 80, 100 ve 120 gün sonra alınmıştır. Daha sonra, alınan embriyolar 0, 0.5 ve 1 mg l⁻¹ GA₃ içeren Murashige ve Tucker (MT) besi ortamında çimlendirilmiştir. Sonuçlar; en yüksek çimlenme oranlarının tozlanmadan 120 gün sonra alınan embriyolarda 1 mg l⁻¹ GA₃ içeren ortamda %100 ile ve 0.5 mg l⁻¹ GA₃ içeren ortamda %95 olduğunu göstermiştir. Tozlanmadan 80 gün sonra alınan embriyoların 0 mg l⁻¹ GA₃ (kontrol) ve 0.5 mg l⁻¹ GA₃ içeren ortamlarda çimlenme olmamıştır. Genel olarak, ortama GA₃ ilavesi kontrole göre çimlenme oranını artırmaktadır. En fazla üç yapraklı bitki oranı %33.33 ile tozlanmadan 100 ve 120 gün sonra alınan ve sırasıyla 0.5 ve 1 mg l⁻¹ GA₃ içeren ortamlardan elde edilmiştir. Sonuçlar, turunçgillerde embriyo kurtarma için en uygun zamanın tozlanmadan sonraki 120 gün olduğunu ve ortamda GA₃ bulunmasının çimlenmeyi artırdığını göstermiştir.

1. Introduction

Citrus aurantium L. that mainly in good compatibility with main citrus types, productive and resistant against calcerous soil, is commonly used as rootstocks in Mediterranean basin where 22% of world citrus production. However, there is an urgency to develop alternative rootstocks resistant to tristeza (CTV) and well adapted to the constraints of the area because of sour orange rootstock is very sensitive against tristeza virus (Ollitrault et al. 1998) and it has sometimes incompatibilities with some lemon cultivars (Tuzcu 1978; Tuzcu et al. 1999).

Although nucellar embryony is genetically valuable in production of homogeneous seedling, it is a big obstacle for breeding of citrus. There is both nucellar embryos and zygotic embryo in polyembryonic citrus seeds. The total number of embryos per seed varies greatly within a tree, as well as among cultivars and there is little consistency within species having nucellar embryony (Ray 2002).

Cultivars that produce nucellar seedlings also occasionally produce zygotic seedlings, but typically monoembryonic cultivars have not been shown to produce nucellar seedlings (Soost and Cameron 1975).

One of the important problems in breeding of citrus, the zygotic embryo in polyembryonic citrus species must compete both nutrients and space with embryos developed from nucellar tissue (Soost and Roose 1996). This negative situation can be eliminated with using embryo rescue technique. Embryo rescue is a very beneficial method for triploid hybrid citrus plants can be produced by interploid crosses followed by *in vitro* embryo rescue techniques to overcome the problem of embryo abortion and recovery of zygotic triploids from polyembryonic seeds (Yi and Deng 1998; Jaskani et al. 2005; Vilorio et al. 2005). The success of embryo rescue depends on the embryo development stage is excised and the composition of the medium used for germination (Jaskani et al. 2005). Genetic structure of embryo and embryo development stage may affect germination capacity of embryos (Vilorio et al. 2005). The addition GA₃ to culture medium positively contributed to the development of embryos (Button and Kochba 1977; Kunitake et al. 1991; Carimi et al. 1998; Das et al. 2000; Wakana et al. 2004; Jaskani et al. 2005). However, GA₃ concentrations in culture medium may give different results depending on the species or variety. Some authors observed that supplementing 0.01 mg l⁻¹ GA₃ to the medium (Ribeiro et al. 2000; Chagas et al. 2003), supplementing 0.1 mg l⁻¹ GA₃ to the medium (Pasqual et al. 1990; Jumin and Nito 1996) and supplementing 2 mg l⁻¹ GA₃ (Gmitter et al. 1990) favored the growth and development of embryos.

The optimal time for embryo rescue vary significantly according to the species and cultivars. Some researcher found that 50 DAP (Chen and Wang 1986; Wang et al. 1999), 80 DAP (Tan et al. 2007), 100 DAP (Wang et al. 1981; Deng et al. 1996; Tusa et al. 1996) 105 DAP (Scarano et al. 2005) and 118 DAP (Chagas et al. 2005) for the optimal time of embryo rescue. Rangan et al. (1969) declared that nucellar embryos had not yet been found in the developing seeds after 120 days pollination.

Trifoliolate is controlled by two dominant gene in citrus and this features is shown in the hybrids of zygotic dominant. Generally, to determine hybrid embryo some additional experiments require such as cytological, flow cytometry, isoenzyme analysis or molecular analysis (Tusa et al. 2002).

In this study, Common mandarin (*Citrus deliciosa* Ten.) and Carrizo citrange [*Citrus sinensis* (L.) Osb. x *Poncirus trifoliata* (L.) Raf.] have been hybridized in order to find new rootstock genotypes. Immature embryos taken 80, 100 and 120 DAP were cultured in MT culture medium supplemented with GA₃ concentrations in order to rescue the zygotic embryos.

2. Materials and Methods

Common mandarin (female) and Carrizo citrange (male) at the "Citrus Genetic Resources Collection" of Bati Akdeniz Agricultural Research Institute were used as parents. The research area is 7.62 km away from sea and its altitude is 9 meters. Its coordinates are 36° 55' 32.40" N and 35° 00' 35.75" E. Soil has salt less-alkali reaction and calcic. Soil structure is sandy-clay- loam as general in soil structure of Antalya region.

Unopened or partially opened flower buds of Carrizo citrange as paternal parent was collected before anthesis and petals and styles were removed with forceps in the laboratory. The flowers were placed at 25°C to promote anther dehiscence. The flowers of Common mandarin as maternal parent were emasculated prior to opening and emasculated flowers were pollinated by touching the dehiscent anthers of the pollen parent gently to the stigma with brush and paper or linen bags were replaced after pollination and were removed approximately one month later. Then fruits were covered by net and marked with labels. Cultural practices like irrigation, fertilizing, weed control, disease and pesticides control were applied on time and according to the technique to the trees.

Modified Murashige and Tucker (1969) medium was used as basic culture medium for embryo germination and 50 g l⁻¹ sucrose, 25 mg adenine sulfate, 500 mg l⁻¹ malt extract were put in medium. Then, 0, 0.5 and 1.0 mg l⁻¹ GA₃ were supplemented to the prepared medium and medium pH was adjusted to 5.7 and then 8 g l⁻¹ agar was added. After sterilization, the prepared medium was distributed 40 ml volume per petri dish.

For embryo culture, hybrid fruits were harvested at 80, 100 and 120 days after cross-pollination. Fruits were washed with water and detergent later, surface-sterilized by immersion for 5 min in 70% ethyl alcohol and 30 min in 20% sodium hypochlorite (Ollitrault et al. 2007). Then, the fruits were cut horizontal with the care of not damaging seeds. The seeds were removed from the fruit by forceps and immature embryos were taken from the microphyl parts of seeds by cutting with a surgical blade under binocular. Two embryos were placed into each petri dish containing modified MT culture medium

Later, the embryos were incubated at 25°C and 90% humidity, in 16 h photoperiod with 1000 lux light intensity in growth room. The embryos which have cotyledon leaves were counted and embryo germination ratio (%) was determined by rating the germinated embryo number to total embryo number.

Germinated embryos were sub-cultured Murashige and Skoog (1962) medium containing 0.02 mg l⁻¹ NAA and 20 g l⁻¹ sucrose in culture tubes to provide seedling growing (Perez-Tornero and Porras 2008). Later, the seedlings were incubated at 25°C and 90% humidity, in 16 h photoperiod with 1000 lux light intensity in growth room.

Trifoliolate seedlings in sub-culture were counted and trifoliolate ratio (%) was determined. Developed plants were transferred into 12x8 cm plastic pots. The seedlings were taken to growth rooms at 25-26°C temperature and 80-85% humidity in order to increase the survival rates. Later, the plants were

transferred to greenhouses for acclimatization to *in vivo* conditions.

The experiment was conducted as random plots with 10 replications and each replications have two embryo. Data were subjected to analysis of variance with mean separation by Duncan's multiple range test. Before comparing the percentage ratio with variance analyses, square root transformation was made to the data.

3. Results

3.1. Germination of embryos

Embryo development stages (day after pollination), GA₃ concentrations in the medium and their interactions were significant on germination rate of Common mandarin x Carrizo citrange hybrid embryos ($p \leq 0.05$).

High germination rates were materialized as 100% and on 95% to taken embryos from 120 DAP in supplemented with 1 mg l⁻¹ GA₃ and 0.5 mg l⁻¹ GA₃ in MT culture medium respectively. The embryos taken from 80 days after pollination did not germinate on MT medium without GA₃ (control) and including with 0.5 mg l⁻¹. The including of 1 mg l⁻¹ GA₃ in medium was increased the ratio of germination of taken embryos at 80, 100 and 120 DAP compared to the control. However, medium supplemented with 0.5 mg l⁻¹ GA₃ did not affect the ratios of germination of taken embryos at 80 DAP. The ratios of germination of taken embryos at 100 DAP decreased while the ratios of germination of taken embryos at 120 DAP increased at medium supplemented with 0.5 mg l⁻¹ GA₃. The germination ratio of the embryos increased with the progress of the embryo development stage (Table 1).

3.2. Trifoliate plant ratio

The highest trifoliate ratio was determined as 33.33% at the embryos taken from 100 and 120 DAP and medium including 0.5 and 1 mg l⁻¹ GA₃ consecutively. Because of the fact that embryos taken from 80 days after pollination did not germinate on MT medium without GA₃ (control) and including with 0.5 mg l⁻¹, trifoliate rate could not determine at these applications. The embryo development stage and the including of GA₃ to the culture medium did not affect the trifoliate rate (Table 2).

4. Discussion and Conclusions

As the highest germination ratio was obtained from embryos taken from 80, 100 and 120 days after pollination at culture medium supplemented with 1 mg l⁻¹ GA₃, this study shown that the suitable dose as 1 mg l⁻¹ GA₃ for immature embryo germination. Our results are in agreement with those of other authors; it have been accepted adding 1 mg l⁻¹ GA₃ to the culture medium for different citrus types and species (Button and Kochba 1977; Kunitake et al. 1991; Carimi et al. 1998; Das et al. 2000; Wakana et al. 2004; Jaskani et al. 2005).

On the other hand, there were some results showing that high embryo germinations were obtained in adding 0.01 mg l⁻¹ GA₃ (Ribeiro et al. 2000; Chagas et al. 2003), 0.1 mg l⁻¹ GA₃ (Pasqual et al. 1990; Jumin and Nito 1996) and 2 mg l⁻¹ GA₃ (Gmitter et al. 1990) in culture medium. These situations may vary due to the embryo development stage, species and cultivars used. There is not an ideal dose for a cultivars.

However, adding 0.01 mg l⁻¹ GA₃ (Ribeiro et al. 2000; Chagas et al. 2003), 0.1 mg l⁻¹ GA₃ (Pasqual et al. 1990; Jumin and Nito 1996) and 2 mg l⁻¹ GA₃ (Gmitter et al. 1990) didn't give good results. This proves that there is not an ideal GA₃ dose for species and varieties and the results may vary according to the species, varieties and embryo development stages.

There is linear relationship between embryo development stage and germination since germination rates from embryos taken 120 DAP at all culture medium was found higher than other. As shown in the results, Carimi et al. (1998) and Vilorio et al. (2005) obtained the best germination ratio in citrus embryos from the embryos taken 120 DAP and stated that genetic structure of embryo and embryo development stage affect germination. On the other hand, there are some results showing that good embryo germinations were obtained at 50 DAP (Chen and Wang 1986; Wang et al. 1999), 80 DAP (Tan et al. 2007), 100 DAP (Wang et al. 1981; Deng et al. 1996; Tusa et al. 1996), 105 DAP (Scarano et al. 2005), and 118 DAP (Chagas et al. 2005) after pollination. These differences may be due to the growing location and cultivars used.

The embryo development stage and the including of GA₃ to the culture medium did not affect the trifoliate rate. It seen that trifoliate rates increased with progressive of embryo development stage on culture medium supplemented with 1 mg l⁻¹ GA₃.

Table 1. The germination rates of embryos Common mandarin x Carrizo citrange obtained from various embryo development stage and different GA₃ doses (%).

Embryo stage	GA ₃ doses			Means of embryo stage
	Control	0.5 mg l ⁻¹	1 mg l ⁻¹	
80 days	0.00 ± 0.00	0.00 ± 0.00	35.00 ± 0.15	11.67 ± 0.06 c*
100 days	65.00 ± 0.13	45.00 ± 0.12	85.00 ± 0.08	65.00 ± 0.07 b
120 days	80.00 ± 0.11	95.00 ± 0.05	100.00 ± 0.00	91.67 ± 0.04 a
Means of GA ₃ doses	48.33 ± 0.08 B	46.67 ± 0.08 B	73.33 ± 0.07 A	

* Mean separation within columns and main effects by Duncan's multiple range test, $p \leq 0.05$. Capital letters indicate doses and small letters indicate embryo taking time.

Table 2. Trifoliate rates of Common mandarin x Carrizo citrange seedling (%).

Trifoliate seedling rates (%)								
Control			0.5 mg l ⁻¹ GA ₃			1 mg l ⁻¹ GA ₃		
80 days	100 days	120 days	80 days	100 days	120 days	80 days	100 days	120 days
0	20.00	30.77	0	33.33	23.53	20.00	21.43	33.33

The results show that it was necessary to supplement GA₃ in the culture medium at citrus according to the species and varieties for embryo development and the appropriate GA₃ dose may vary according to the species and varieties. However, as obtained germination and trifoliate rates is higher on 120 days prove that the optimal time for embryo rescue at citrus is 120 days after pollination.

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References

- Button J, Kochba J (1977) Tissue culture in the citrus industry. In: Reinert J, Bajaj YPS (eds.). Applied and Fundamental Aspects of Plant Cell, Tissue and Organ Culture, Springer-Verlag. pp. 70-92.
- Carimi F, Pasquale F, Puglia AM (1998) *In vitro* rescue of zygotic embryos of sour orange, *Citrus aurantium* L., and their detection based on RFLP analysis. Plant Breeding 117: 261-266.
- Chagas EA, Pasqual M, Ramos JD, Cardoso P, Cazetta JO, Figueiredo MAD (2003) Development of globular embryos from the hybridization between 'Pera Rio' sweet orange and 'Ponca' mandarin. Rev Bras Frutic Jaboticabal-SP 25(3): 483-488.
- Chagas E, Pasqual AM, Ramos JD, Pio LAS, Dutra LF, Cazetta JO (2005) Activated charcoal and gibberellic acid concentrations on immature embryos culture. Ciencia e Agrotecnologia Lavras: Universidade Federal de Lavras 29(6): 1125-1131.
- Chen ZG, Wang JF (1986) Plantlets derived from early *in vitro* culture of citrus zygotic embryo. Journal of Fujian Agricultural and Forestry University 15(4): 271-276.
- Das A, Paul AK, Chaudhuri S (2000) Micro propagation of sweet orange *Citrus sinensis* Osbeck for the development of nucellar seedling. Indian Journal of Experimental Biology 38(3): 269-272.
- Deng XX, Yi HL, Li F, Guo WW (1996) Triploid plants regenerated from crossing diploid pumelo and tangerine with allotetraploid somatic hybrid of citrus. Proceedings of International Society Citriculture 1: 189-192.
- Gmitter FG, Ling XB, Deng XX (1990) Induction of triploid *Citrus* plants from endosperm calli *in vitro*. Theoretical and Applied Genetics 80: 785-790.
- Jaskani MJ, Khan IA, Khan MM (2005) Fruit set, seed development and embryo germination in interploid crosses of citrus. Scientia Horticulturae 107: 51-57.
- Jumin HB, Nito N (1996) Plant regeneration via somatic embryogenesis from protoplast of six plant species related to *Citrus*. Plant Cell Reports 15(5): 332-336.
- Kunitake H, Kagami H, Mii M (1991) Somatic embryogenesis and plant regeneration from protoplasts of 'Satsuma' mandarin (*Citrus unshiu* Marc.). Scientia Horticulturae 47: 27-33.
- Murashige T, Skoog F (1962) A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiologia Plantarum 15: 473-497.
- Murashige T, Tucker PH (1969) Growth factor requirements of citrus tissue culture. Proceeding of 1st International Citrus Symposium 3: 1155-1161.
- Ollitrault P, Dambier D, Froelicher Y, Seker M (1998) Rootstock breeding strategies for the Mediterranean citrus industry: The somatic hybridization potential. Acta Horticulturae 535: 157-162.
- Ollitrault P, Guo W, Grosser JW (2007) Somatic hybridization, citrus genetics, breeding and biotechnology. In: Khan IA (eds.). Citrus genetics, breeding and biotechnology. Institute of Horticultural Sciences, University of Agricultural Faisalabad, 38040, Pakistan, pp. 235-260.
- Pasqual M, Riberio VG, Ramos JD (1990). Influencia do GA₃ e do carvaoativa dosobre o enraizamento *in vitro* de embriões de laranja 'Natal'. Pesquisa Agropecuaria Brasileira, Brasilia 25(10): 1477-1482.
- Perez-tornero O, Porras I (2008) Assessment of polyembryony in lemon: rescue and *in vitro* culture of immature embryos. Plant Cell Tissue Organ Culture 93: 173-180.
- Rangan TS, Murashige T, Bitters WP (1969) *In vitro* study of zygotic and nucellar embryogenesis in citrus. Proceedings of International Society Citriculture 1: 225-229.
- Ray PK (2002) Breeding tropical and subtropical fruits. Springer-Verlag, Berlin, Heidelberg, pp. 96-98.
- Ribeiro VG, Sanábio D, de Souza CN, Lopes PSN, Bocardo MR, Pasqual M (2000) Efeitos de ácidogiberélico e carvãoativado no cultivo *in vitro* de *Citrus limonia* Osbeck x *Poncirus trifoliata* (L.) Raf. Pesquisa Agropecuaria Brasileira, Brasilia 35: 27-30.
- Scarano MT, Tusa N, Abbate L, Lucretti S, Nardi L, Ferrante S (2005) Flow cytometry, SSR and modified AFLP markers for the identification of zygotic plantlets in backcrosses between 'Femminello' lemon hybrids (2n and 4n) and a diploid clone of 'Femminello' lemon (*Citrus limon* L. Burm. F.) tolerant to *mal secco* disease. Plant Science 164(6): 1009-1017.
- Soost RK, Cameron JW (1975) Citrus. In: Janick J, Moore JN (Eds.), Advances in fruit breeding. Purdue University Press, West Lafayette, pp. 507-540.
- Soost RK, Roose M (1996) Citrus. 1: 257-323. In: Jules J, Moore JN (eds). Fruit breeding: Tree and tropical fruits, John Wiley and Sons Inc., New York.
- Tan M, Song J, Deng X (2007) Production of two mandarin x trifoliate orange hybrid populations via embryo rescue with verification by SSR analysis. Euphytica 157: 155-160.
- Tusa N, Fatta del Bosco S, Nardi L, Lucretti S (1996) Obtaining triploid plants by crossing citrus lemon cv. 'Femminello' 2N x 4N allotetraploid somatic hybrids. Proceedings of International Society Citriculture 1: 133-136.
- Tusa N, Abbate L, Ferrante S, Lucretti S, Scarano MT (2002) Identification of zygotic and nucellar seedlings in *Citrus* interploid crosses by means of isozymes, flow cytometry, and ISSR-PCR. Cellular and Molecular Biology Letters 7: 703-708.
- Tuzcu O (1978) Turunçgillerde anaç ve sorunları. Çağdaş Tarım Tekniği 3: 31-35.
- Tuzcu O, Yıldırım B, Düzenoglu S, Bahceci I (1999) Değişik turunçgil anaçlarının Washington Navel ve Moro kan portakal çeşitlerinin verim ve kalitesi üzerine etkileri. Turkish Journal of Agriculture and Forestry 23: 213-222.
- Viloria Z, Grosser JW, Bracho B (2005) Immature embryo rescue, culture and seedling development of acid citrus fruit derived from interploid hybridization. Plant Cell, Tissue and Organ Culture 82: 159-167.
- Wakana A, Binh XN, Iwamasa M (2004) Germinability of embryos during seed development in *Citrus* (Rutaceae). Journal of the Faculty of Agriculture, Kyushu University Fukuoka: Faculty of Agriculture Publication, Kyushu University 49(1): 49-59. (In Japanese with English abstract).
- Wang YS, Zhou BY, Gao SX (1981) The study on embryo culture of citrus (II) Early *in vitro* culture of immature embryos of polyembryonic cultivars. Acta Horticulturae 8(1): 13-17.
- Wang JF, Chen ZG, Lin TX (1999) Observation on the embryonic development in *citrus* after cross pollination. Chinese Development. Society for Reproductive Biology 8(2): 57-63.
- Yi HL, Deng XX (1998) A study of culture of citrus triploid plantlets. Journal of Fruit Science 15(3): 212-216.